NAME: OLUPINYO TOSIN ELIZABETH

MATRIC NUMBER: 16/MHS06/055

COURSE CODE: MLS 201 (GLASSWEAR CLEANING)

QUESTION: DESCRIBE THE CLEANING METHODS FOR 4 NAMED LABORATORY GLASSWEARS.

1. **Culture Test Tubes:** Used culture tubes need to be sterilized prior to cleaning. The most dependable technique for sterilizing culture tubes is to autoclave it for half an hour at around 121°C (15 p.s.i. pressure). As media gets solidified on cooling, it should be poured out at the time the tubes are still hot. The tubes when emptied, must be brushed with water and detergent and then rinsed with running tap water. After this it should be rinsed with distilled water, and placed properly for drying. When tubes are filled with media that is sterilized by autoclaving, there is no need to plug until the addition of the media. Thu media and tubes are both sterilized with one step autoclaving.
2. **Cleaning PYREX Fritted Ware:** A new fritted filter should be washed by suction with hot hydrochloric acid and then rinsed with water before it is used. This treatment will remove loose particles of foreign matter such as dust. It is advisable to clean all PYREX fritted filters as soon as possible after use. This will prolong their life. Many precipitates can be removed from the filter surface simply by rinsing from the reverse side with water under pressure not exceeding 15 lb/sq. in. Drawing water through the filter from the reverse side with a vacuum pump is also effective. Some precipitates tend to clog the pores of a fritted filter and may require special cleaning solutions.
3. **Cleaning Corning Slides and Cover Glass**: It is especially important that microscope slides and cover glass used for the preparation of blood films, bacteriologic smears or cell culture be perfectly clean and free from scratches. Slides should be washed, placed in glacial acetic acid for 10 minutes, rinsed with distilled water and wiped dry with clean paper towels or cloth. Before use, wash with alcohol and wipe dry. Or the slides, after acid treatment and rinsing, may be placed in a wide jar and covered with alcohol. As needed, remove from the jar and wipe dry.
4. **Cleaning PYREX Blood Chemistry Pipets**: After use, rinse thoroughly with cool tap water, distilled water, alcohol or acetone, and then either. Dry by suction. (Do not blow into the pipets as this will cause moisture to condense on the inside of the pipet). To remove particles of coagulated blood or dirt, a cleaning solution should be used. One type of solution will suffice in one case, whereas a stronger solution may be required in another. It is best to fill the pipet with the cleaning solution and allow it to stand overnight. Sodium hypochlorite (laundry bleach) or a detergent may be used. Hydrogen peroxide is also useful. In difficult cases, use concentrated nitric acid. Some particles may require loosening with a horse hair or piece of fine wire. Take care not to scratch the inside of the pipet