**NAME: OLOKOR TOBORE DAVID**

**MATRIC NO: 16/MHS06/052**

**DEPARTMENT: MEDICAL LAB SCIENCE**

**COURSE: MLS 201**

**ASSIGNMENT**

Describe the cleaning methods for 4 named laboratory glasswares.

**ANSWERS**

* **Glassware Used**[**for Organic Chemistry**](https://www.thoughtco.com/definition-of-organic-chemistry-604593):

Rinse the glassware with the appropriate solvent. Use deionized water for water-soluble contents. Use ethanol for ethanol-soluble contents, followed by rinses in deionized water. Rinse with other solvents as needed, followed by ethanol and finally deionized water. If the glassware requires scrubbing, scrub with a brush using hot soapy water, rinse thoroughly with tap water, followed by rinses with deionized water.

* **Burettes:**

Wash with hot soapy water, rinse thoroughly with tap water, and then rinse 3-4 times with deionized water. Be sure the final rinses sheet off of the glass. Burettes need to be thoroughly clean to be used for quantitative lab work.

* **Pipettes and**[**Volumetric Flasks**](https://www.thoughtco.com/definition-of-volumetric-flask-605783):

In some cases, you may need to soak the glassware overnight in soapy water. Clean pipettes and [volumetric flasks](https://www.thoughtco.com/what-is-a-volumetric-flask-606043) using warm soapy water. The glassware may require scrubbing with a brush. Rinse with tap water followed by 3-4 rinses with deionized water.

**Culture Tubes**

* Culture tubes which have been used previously must be sterilized before cleaning. The best general method for sterilizing culture tubes is by autoclaving for 30 minutes at 121°C (15ib. pressure). Media which solidify on cooling should be poured out while the tubes are emptied, brush with detergent and water, rinse thoroughly with tap water, rinse with distilled water, place in a basket and dry, sterile container. It may be expedient to sterilize all tubes as routine.
* To clean and sterilize tubes containing blood, discard the clots in a waste container and place the tubes in a large basket. Put the basket, with others, in a large bucket or boiler. Cover with water, add a fair quantity of soap or detergent and boil for 30 minutes. Rinse the tubes and clean with brush, rinse and
* dry with the usual precautions.  
  It is imperative when washing serological glassware that all acid, alkali and detergent be completely removed, Both acid and alkali in small amounts destroy complement and in large amounts produce hemolysis. Detergents interfere with s e r o I o g i c reactions.  
  Serological tubes and glassware should be kept separate from all other glassware and used for nothing except serologic procedures.