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COURSE; MLS201 (GLASSWARE CLEANING)

ASSIGINMENT

Describe the cleaning methods of four laboratory equipment.

1. **CLEANING METHOD FOR BURRET-**
2. Rinse with distilled water:
With the stopcock closed, add some distilled water to the burette. Tip and roll the burette, allowing the water to have contact with the entire inside surfaces. Open the stopcock and allow the water to drain. If the water drains without leaving any droplets on the side, repeat the rinse twice more then move to step two. If droplets remain on the inside surface, wash the burette with detergent solution, rinse several times with tap water and then rinse three times with distilled water.
3. Rinse with solution:
After draining the final distilled water rinse, close the stopcock and add about 5 mL of the solution to be dispensed from the burette. Again, roll and tip the burette so the solution has contact with all the inside surfaces. Open the stopcock and allow the solution to drain. Repeat this twice more. Discard the solution used in the rinses.
4. After you are finished with the burette in your experiment, rinse it by filling it with distilled water and allowing it to drain.
5. **CLEANING METHOD FOR BEAKERS-**
* For water soluble solutions;

(e.g., sodium chloride or sucrose solutions) Rinse 3-4 times with deionized water then put the beaker away.

* For Water insoluble solutions;

(e.g., solutions in hexane or chloroform) Rinse 2-3 times with ethanol or acetone, rinse 3-4 times with deionized water, then put the beaker away. In some situations, other solvents need to be used for the initial rinse.

* For Strong acids;
(e.g., concentrated HCl or H2SO4) Under the fume hood, carefully rinse the beaker with copious volumes of tap water. Rinse 3-4 times with deionized water, then put the beaker away.
* For Strong bases;
(e.g., 6M NaOH or concentrated NH4OH) Under the fume hood, carefully rinse the beaker with copious volumes of tap water. Rinse 3-4 times with deionized water, then put the beaker away.
* For weak acids;
(e.g., acetic acid solutions or dilutions of strong acids such as 0.1M or 1M HCl or H2SO4) Rinse 3-4 times with deionized water before putting the beaker away.
* For Weak bases;

 (e.g., 0.1M and 1M NaOH and NH4OH) Rinse thoroughly with tap water to remove the base and then rinse 3-4 times with deionized water before putting the beaker away.

1. **CLEANING METHODS FOR MICROSCPE SLIDES-**

Place all dirty microscope slides in a water basin full of warm water and detergent. Place them in the basin carefully so that none of them touch. Leave the slides in the basin for an entire day. This should be sufficient time to allow the blood, oil or other material to loosen. Use gauze to rub each slide individually on both sides until they are clean. Leave the slides in the water for a few days at the most, leaving the slides in water with detergent in it for weeks or longer risk letting the water evaporate. This will leave a detergent residue on the slides that will be difficult to remove. Wrap cleaned slides in sheets of clean paper until they are ready to be used again. This allows storing of the slides closer together if a case isn’t available. Store slides somewhere dry to avoid the slides sticking together due to humidity.

1. **CLEANING METHODS FOR PETRI DISHES-**
* Remove the media from the petri dish
* Immerse the plates in 1%v/v (1ml in 100ml) soap solution for 30minutes
* Wash the petri plates with plenty of water. Rinse with freshly distilled water
* Sterilize the petri plates as per SOP for sterilization of glassware
* Store the petri plates properly.